



International Journal of Innovative Technologies in Social Science

e-ISSN: 2544-9435

Scholarly Publisher
RS Global Sp. z O.O.
ISNI: 0000 0004 8495 2390

Dolna 17, Warsaw,
Poland 00-773
+48 226 0 227 03
editorial_office@rsglobal.pl

ARTICLE TITLE

ROLE OF HIF-1 SIGNALING PATHWAY IN CELLULAR
ADAPTATION TO HYPOXIA

DOI

[https://doi.org/10.31435/ijitss.4\(48\).2025.4299](https://doi.org/10.31435/ijitss.4(48).2025.4299)

RECEIVED

23 October 2025

ACCEPTED

25 December 2025

PUBLISHED

30 December 2025

LICENSE



The article is licensed under a **Creative Commons Attribution 4.0 International License**.

© The author(s) 2025.

This article is published as open access under the Creative Commons Attribution 4.0 International License (CC BY 4.0), allowing the author to retain copyright. The CC BY 4.0 License permits the content to be copied, adapted, displayed, distributed, republished, or reused for any purpose, including adaptation and commercial use, as long as proper attribution is provided.

ROLE OF HIF-1 SIGNALING PATHWAY IN CELLULAR ADAPTATION TO HYPOXIA

Kamil Borysewicz (Corresponding Author, Email: kamilborys.borysewicz@gmail.com)

The Municipal Specialist Hospital in Toruń, Toruń, Poland

ORCID ID: 0000-0003-4510-2759

Illia Koval

Provincial Hospital in Poznań, Poznań, Poland

ORCID ID: 0009-0002-1109-6354

Wiktor Kubik

5th Military Clinical Hospital in Krakow, Krakow, Poland

ORCID ID: 0009-0000-4041-0846

Bartłomiej Czarnecki

Provincial Specialist Hospital No. 5 named after St. Barbara in Sosnowiec, Sosnowiec, Poland

ORCID ID: 0009-0006-8960-5760

Jan Nowak

Dr. Emil Warmiński Clinical Hospital of the Bydgoszcz University of Technology – Independent Public Health Care Facility, Bydgoszcz, Poland

ORCID ID: 0009-0006-8145-8647

Barbara Kujawa

Provincial Hospital in Poznań, Poznań, Poland

ORCID ID: 0009-0000-3951-965X

Bartosz Zwoliński

Central Clinical Hospital of the Medical University in Warsaw, Warsaw, Poland

ORCID ID: 0009-0000-8675-3828

Kacper Sukiennicki

District Hospital in Chrzanów, Chrzanów, Poland

ORCID ID: 0009-0003-6864-4996

Wirginia Bertman

Stefan Żeromski Specialist Hospital, Kraków, Poland

ORCID ID: 0009-0002-9166-8681

Natalia Koldej

District Hospital in Ilża, Ilża, Poland

ORCID ID: 0009-0004-3203-8019

Zuzanna Kępczyńska

St. Anne's Hospital in Piaseczno, Piaseczno, Poland

ORCID ID: 0009-0005-2360-854X

Katarzyna Szewczyk

L. Rydygier Specialist Hospital in Krakow, Krakow, Poland

ORCID ID: 0009-0008-7451-3091

Klaudia Romejko

Provincial Hospital in Poznań, Poznań, Poland

ORCID ID: 0009-0003-6452-1323

ABSTRACT

Hypoxia accompanying chronic inflammatory diseases leads to dysregulation of cells' homeostasis, causes energy deficits and intensifies inflammatory processes. Hypoxia-inducible factors (HIFs) are the central regulators of the response to hypoxia, enabling metabolic and functional adaptation of immune cells by altering gene expression. HIF-1 α and HIF-2 α modulate the lifespan, differentiation, priming and activation of neutrophils, macrophages, lymphocytes, and dendritic cells, playing a key role in influencing the balance between proinflammatory and reparative responses. Interactions between HIF and NF- κ B pathways regulate hypoxic and inflammatory signaling, and as a result determine course, severity and treatment efficacy of many chronic diseases. Those two pathways are capable of influencing each other in a manner of negative feedback-loop. Understanding the mechanisms of this regulation opens up new therapeutic perspectives in the treatment of pulmonary diseases associated with hypoxia and inflammation, and a hypothetical possibility to slow down the ratio at which chronic inflammatory response contributes to deterioration of patients' quality of life.

KEYWORDS

Hypoxia-inducible Factor; HIF Signaling Pathway; NF- κ B Signaling Pathway, Cellular Adaptation to Hypoxia, Inflammatory Response

CITATION

Kamil Borysewicz, Illia Koval, Wiktor Kubik, Bartłomiej Czarnecki, Jan Nowak, Barbara Kujawa, Bartosz Zwoliński, Kacper Sukiennicki, Wirginia Bertman, Natalia Kołdej, Zuzanna Kępczyńska, Katarzyna Szewczyk, Klaudia Romejko. (2025) Role of HIF-1 Signaling Pathway in Cellular Adaptation to Hypoxia. *International Journal of Innovative Technologies in Social Science*. 4(48). doi: 10.31435/ijitss.4(48).2025.4299

COPYRIGHT

© The author(s) 2025. This article is published as open access under the **Creative Commons Attribution 4.0 International License (CC BY 4.0)**, allowing the author to retain copyright. The CC BY 4.0 License permits the content to be copied, adapted, displayed, distributed, republished, or reused for any purpose, including adaptation and commercial use, as long as proper attribution is provided.

Introduction

In numerous pathological conditions, including chronic inflammatory lung diseases, pulmonary fibrosis, and chronic respiratory failure, there is an imbalance between the oxygen demand of cells and its supply [1-3]. A deficit of oxygen, which is the final electron acceptor in the mitochondrial respiratory chain, leads to collapse of mitochondrial bioenergetic function, reducing yield of adenosine triphosphate (AT) production, and in consequence insufficient energy for cell's physiological processes [2]. In areas of ongoing inflammation, a specific hypoxic microenvironment develops, referred to as "inflammatory hypoxia" [3]. Accelerated metabolism of activated immune cells (neutrophils, monocytes) promotes cell proliferation and the expression of oxygenase enzymes, which further deepens deficit of oxygen [4]. By exacerbating tissue dysfunction pathological hypoxia contributes to disease's progress through the dysregulation of the immune response [5]. Furthermore, state of hypoxia by itself influences the course of inflammatory processes by regulating the activity of oxygen-dependent signaling pathways in many types of immune cells [6,7].

HIF Pathway Modulates Cellular Adaptation in Response to Hypoxia

HIF transcriptional complex acts as the key regulator of the cellular response to oxygen deficiency [8]. The discovery of the molecular mechanism of HIF-dependent cellular adaptation as a response to hypoxia was awarded the Nobel Prize in Physiology or Medicine in 2019 (W.G. Kaelin, G.L. Semenza, P.J. Ratcliffe) [9].

HIF is a heterodimeric transcription factor consisting of an oxygen-regulated HIF- α subunit and a constitutively present HIF-1 β subunit, also known as aryl hydrocarbon receptor nuclear translocator (ARNT). So far, three isoforms of the HIF- α have been identified: HIF-1 α ; HIF-2 α ; and HIF-3 α , each differing in the range of regulated genes and their function [10-12]. **Table 1.** includes key differences between HIF- α isoforms.

Table 1. Isoforms of HIF- α

Isoform	Primary Functions	Clinical significance
HIF-1 α	Regulation of glycolysis, cell survival in hypoxia	Inflammation Targeting Cancers
HIF-2 α	Angiogenesis, endothelial cell proliferation	Anemia Targeting Cancers
HIF-3 α	Regulation of HIF-1/2 activity, less known	Potential biomarker

In conditions of normoxia, HIF- α is hydroxylated by the dioxygenase families (PHD1-3 and FIH), then degraded by the von Hippel-Lindau protein (pVHL) via 26S proteasome [13]. Hypoxia inhibits the activity of hydroxylases, leading to the stabilization of HIF- α , its dimerization with HIF-1 β , and activation of target genes through binding to HRE (hypoxia response element) sequences in DNA [14].

HIF, NF- κ B and Intertwinement of Two Signaling Pathways

HIF-signal pathway interacts strongly with another central pathway regulating the inflammatory response, the nuclear transcription factor NF- κ B [15,16]. Both pathways share regulators, target genes and activate in response to cytokines and pathogens. PHD hydroxylases, which control HIF stability, also participate in regulation of NF- κ B pathway [17].

In conditions of chronic inflammation, typical for example in COPD, increased NF- κ B activity enhances HIF-1 α transcription, which promotes hypoxia [18].

Studies in animal models of COPD have shown that pharmacological inhibition of NF- κ B activity reduces HIF-1 α expression, leading to a reduction in inflammation and improvements in epithelium [19].

HIF Signaling Pathway Regulates Immune Cells' Metabolism

HIF controls the effector function of immune cells primarily by regulating their metabolism [14,25–27], as per **Table 2**.

Table 2. Effect of HIF on various immune cells.

Cell Type	Effect of HIF	Results
Neutrophils	Extended Lifespan; Production of antimicrobial peptides	Innate immunity Increase
Macrophages	M1/M2 polarization; migration to inflammatory foci	Increase of Inflammatory Response
T cells	Th17/Treg equilibrium control; CD8 ⁺ cytotoxic activity	Autoimmunity, Anti-Cancer Cells
B Lymphocytes	Proliferation, Promotion of humoral response	Increase in Antibody Production
Mast cells/eosinophils	Production of Cytokines, VEGF	Asthma, allergic reactions

In inflammatory niches, cells migrate from the blood to an environment with low pO₂ and must undergo metabolic adaptation. In granulocytes, glycolysis—regulated by HIF—is the main source of ATP [26]. Both isoforms (HIF-1 α and HIF-2 α) reduce dependence on OXPHOS, inhibit the TCA cycle, and shift metabolism toward anaerobic ATP production [28–30].

HIF-1 α prolongs neutrophils' lifespan, increases the expression of β 2-integrins, the production of antibacterial peptides and key glycolysis enzymes; the absence of HIF-1 α impairs ATP generation, aggregation, chemotaxis and bacterial killing [26,31,32].

Macrophages- HIF-1 α promotes macrophages' differentiation into M1 (pro-inflammatory) phenotype, while HIF-2 α promotes the M2 (repair) phenotype. Deletion of HIF-1 α reduces ATP, survival, invasion, and bactericidal activity [32]; HIF-1 α increases migration to sites of infection via \uparrow CXCR4 and \downarrow CCR5 [14]. IL-8 is a target of HIF-1 α ; increased secretion of IL-8 by alveolar macrophages may accelerate ARDS.

Dendritic Cells- HIF-dependent metabolic reprogramming participates in dendritic cells' activation, differentiation, proliferation, migration, and apoptosis; HIF regulates the production of IFN- γ , IL-22, IL-10, and the expression of chemokine receptors that attract neutrophils [14,31,33].

T lymphocytes - HIF-1 α is crucial for T-cell survival; it controls the Th17/Treg switch (important in autoimmunity) [14,31]. In CD8⁺ T cells, HIF-1 α promotes glycolysis necessary for effector function and the generation of populations with antitumor properties; conditional VHL knockout (permanently stable HIF) accelerates the differentiation of long-lived memory effectors against viral infections. HIF-1 α deletion limits CTL infiltration and their killing of cancer cells [25]. Hypoxia and HIF enhance glycolysis in B cells, affecting the development, proliferation, apoptosis, and maturation of high-affinity antibody responses [14].

Mast cells, eosinophils, basophils. HIF-1 α stabilization supports survival and function; after TLR stimulation, HIF-1 α enhances IL-8 and TNF- α and the expression of histidine decarboxylase (\rightarrow histamine) [14]. In the bronchial epithelium, HIF-1 induces VEGF, increasing vascular permeability and airway edema [14]. HIF-1 α and HIF-2 α modulate eosinophil chemotaxis and asthma pathogenesis; HIF-1 α supports basophil activity in chronic inflammation. HIF is also associated with the formation of extracellular traps (METs, NETs) with antimicrobial significance.

Mitochondrial regulation and the role of ROS

Mitochondria participate in the modulation of HIF stability through reactive oxygen species (ROS) generated in the respiratory chain [28]. ROS inhibit the degradation of HIF-1 α and enhance its transcriptional activity. The use of antioxidants such as ebselen reduces HIF-1 α stabilization, confirming the role of oxidative stress in the regulation of this pathway [29].

Clinical Significance and Application

The HIF pathway is an attractive therapeutic target in diseases involving hypoxia and chronic inflammation. The best-known group of substances are HIF hydroxylase inhibitors, which stabilize HIF- α and enhance the adaptive response [30]. Preclinical studies have shown that modulation of HIF activity can both reduce tissue damage and increase antimicrobial resistance [31,32].

At the same time, due to the strong connection between the HIF pathway and other regulatory systems (NF- κ B, p53, Notch), therapeutic targeting requires particular caution and further research [33].

Conclusions

Hypoxia-inducible factors (HIFs) play a central role in regulating gene expression under hypoxic conditions. Stabilization of HIF-1 α enables cells to adapt metabolically and functionally, but may also contribute to immune dysfunction and the progression of chronic diseases. The interaction between HIF and NF- κ B pathways forms the basis for the integration of inflammatory and hypoxic responses, opening up new therapeutic perspectives in the treatment of lung diseases and other conditions associated with chronic inflammation and hypoxia.

Authors' Contributions:

Ilia Koval-methodology; formal analysis

Kamil Borysewicz (corresponding author)— conceptualization; investigation; writing - original draft

Bartłomiej Czarnecki — methodology; formal analysis; project administration

Jan Nowak - methodology; formal analysis

Bartosz Zwoliński — software; validation/check; data curation

Kacper Sukiennicki — resources; writing - original draft

Wirginia Bertman -investigation; writing - review & editing

Natalia Kołdej — investigation; writing - review & editing

Klaudia Romejko — investigation; writing - original draft

Wiktoria Kubik— data curation; writing - original draft

Zuzanna Kępczyńska — data curation; supervision

Katarzyna Szewczyk - validation/check; supervision; project administration

Barbara Kujawa- investigation; writing—review, language check

Conflicts of Interest: No conflicts of interest to declare.

REFERENCES

1. Choudhry, H., & Harris, A. L. (2018). Advances in Hypoxia-Inducible Factor Biology. *Cell metabolism*, 27(2), 281–298. <https://doi.org/10.1016/j.cmet.2017.10.005>
2. Semenza G. L. (2014). Oxygen sensing, hypoxia-inducible factors, and disease pathophysiology. *Annual review of pathology*, 9, 47–71. <https://doi.org/10.1146/annurev-pathol-012513-104720>
3. Bartels, K., Grenz, A., & Eltzschig, H. K. (2013). Hypoxia and inflammation are two sides of the same coin. *Proceedings of the National Academy of Sciences of the United States of America*, 110(46), 18351–18352. <https://doi.org/10.1073/pnas.1318345110>
4. Palazon, A., Goldrath, A. W., Nizet, V., & Johnson, R. S. (2014). HIF transcription factors, inflammation, and immunity. *Immunity*, 41(4), 518–528. <https://doi.org/10.1016/j.immuni.2014.09.008>
5. Taylor, C. T., & Colgan, S. P. (2017). Regulation of immunity and inflammation by hypoxia in immunological niches. *Nature reviews. Immunology*, 17(12), 774–785. <https://doi.org/10.1038/nri.2017.103>
6. Dvornikova, K. A., Platonova, O. N., & Bystrova, E. Y. (2023). Hypoxia and Intestinal Inflammation: Common Molecular Mechanisms and Signaling Pathways. *International journal of molecular sciences*, 24(3), 2425. <https://doi.org/10.3390/ijms24032425>
7. Sitkovsky, M., & Lukashev, D. (2005). Regulation of immune cells by local-tissue oxygen tension: HIF1 alpha and adenosine receptors. *Nature reviews. Immunology*, 5(9), 712–721. <https://doi.org/10.1038/nri1685>
8. Kaelin, W. G., Jr, & Ratcliffe, P. J. (2008). Oxygen sensing by metazoans: the central role of the HIF hydroxylase pathway. *Molecular cell*, 30(4), 393–402. <https://doi.org/10.1016/j.molcel.2008.04.009>
9. Nobel Prize in Physiology or Medicine 2019. The Nobel Assembly, Karolinska Institutet.
10. Keith, B., Johnson, R. S., & Simon, M. C. (2011). HIF1 α and HIF2 α : sibling rivalry in hypoxic tumour growth and progression. *Nature reviews. Cancer*, 12(1), 9–22. <https://doi.org/10.1038/nrc3183>
11. Semenza G. L. (2012). Hypoxia-inducible factors in physiology and medicine. *Cell*, 148(3), 399–408. <https://doi.org/10.1016/j.cell.2012.01.021>
12. Corrado, C., & Fontana, S. (2020). Hypoxia and HIF Signaling: One Axis with Divergent Effects. *International Journal of Molecular Sciences*, 21(16), 5611. <https://doi.org/10.3390/ijms21165611>
13. Schofield, C. J., & Ratcliffe, P. J. (2004). Oxygen sensing by HIF hydroxylases. *Nature reviews. Molecular cell biology*, 5(5), 343–354. <https://doi.org/10.1038/nrm1366>
14. Wenger, R. H., Stiehl, D. P., & Camenisch, G. (2005). Integration of oxygen signaling at the consensus HRE. *Science's STKE : signal transduction knowledge environment*, 2005(306), re12. <https://doi.org/10.1126/stke.3062005re12>
15. Rius, J., Guma, M., Schachtrup, C., Akassoglou, K., Zinkernagel, A. S., Nizet, V., Johnson, R. S., Haddad, G. G., & Karin, M. (2008). NF-kappaB links innate immunity to the hypoxic response through transcriptional regulation of HIF-1alpha. *Nature*, 453(7196), 807–811. <https://doi.org/10.1038/nature06905>
16. van Uden, P., Kenneth, N. S., & Rocha, S. (2008). Regulation of hypoxia-inducible factor-1alpha by NF-kappaB. *The Biochemical journal*, 412(3), 477–484. <https://doi.org/10.1042/BJ20080476>
17. Cummins, E. P., Berra, E., Comerford, K. M., Ginouves, A., Fitzgerald, K. T., Seeballuck, F., Godson, C., Nielsen, J. E., Moynagh, P., Pouyssegur, J., & Taylor, C. T. (2006). Prolyl hydroxylase-1 negatively regulates IkkappaB kinase-beta, giving insight into hypoxia-induced NFkappaB activity. *Proceedings of the National Academy of Sciences of the United States of America*, 103(48), 18154–18159. <https://doi.org/10.1073/pnas.0602235103>
18. Barnes P. J. (2016). Inflammatory mechanisms in patients with chronic obstructive pulmonary disease. *The Journal of allergy and clinical immunology*, 138(1), 16–27. <https://doi.org/10.1016/j.jaci.2016.05.011>
19. Titova, O. N., Kuzubova, O. N., Lebedeva, E. S., Surkova, E. A., Preobrazhenskaya, T. N., & Dvorakovskaya, I. V. (2018). Anti-inflammatory and regenerative effects of hypoxic signaling inhibition in a model of COPD. *PULMONOLOGIYA*, 28(2), 169–176..
20. Cramer, T., Yamanishi, Y., Clausen, B. E., Förster, I., Pawlinski, R., Mackman, N., Haase, V. H., Jaenisch, R., Corr, M., Nizet, V., Firestein, G. S., Gerber, H. P., Ferrara, N., & Johnson, R. S. (2003). HIF-1alpha is essential for myeloid cell-mediated inflammation. *Cell*, 112(5), 645–657. [https://doi.org/10.1016/s0092-8674\(03\)00154-5](https://doi.org/10.1016/s0092-8674(03)00154-5)
21. Imtiyaz, H. Z., & Simon, M. C. (2010). Hypoxia-inducible factors as essential regulators of inflammation. *Current topics in microbiology and immunology*, 345, 105–120. https://doi.org/10.1007/82_2010_74
22. Corzo, C. A., Condamine, T., Lu, L., Cotter, M. J., Youn, J. I., Cheng, P., Cho, H. I., Celis, E., Quiceno, D. G., Padhya, T., McCaffrey, T. V., McCaffrey, J. C., & Gaborilovich, D. I. (2010). HIF-1 α regulates function and differentiation of myeloid-derived suppressor cells in the tumor microenvironment. *The Journal of experimental medicine*, 207(11), 2439–2453. <https://doi.org/10.1084/jem.20100587>
23. Walmsley, S. R., Print, C., Farahi, N., Peyssonnaud, C., Johnson, R. S., Cramer, T., Sobolewski, A., Condliffe, A. M., Cowburn, A. S., Johnson, N., & Chilvers, E. R. (2005). Hypoxia-induced neutrophil survival is mediated by HIF-1alpha-dependent NF-kappaB activity. *The Journal of experimental medicine*, 201(1), 105–115. <https://doi.org/10.1084/jem.20040624>

24. Tannahill, G. M., Curtis, A. M., Adamik, J., Palsson-McDermott, E. M., McGettrick, A. F., Goel, G., Frezza, C., Bernard, N. J., Kelly, B., Foley, N. H., Zheng, L., Gardet, A., Tong, Z., Jany, S. S., Corr, S. C., Haneklaus, M., Caffrey, B. E., Pierce, K., Walmsley, S., Beasley, F. C., ... O'Neill, L. A. (2013). Succinate is an inflammatory signal that induces IL-1 β through HIF-1 α . *Nature*, 496(7444), 238–242. <https://doi.org/10.1038/nature11986>
25. Dang, E. V., Barbi, J., Yang, H. Y., Jinasena, D., Yu, H., Zheng, Y., Bordman, Z., Fu, J., Kim, Y., Yen, H. R., Luo, W., Zeller, K., Shimoda, L., Topalian, S. L., Semenza, G. L., Dang, C. V., Pardoll, D. M., & Pan, F. (2011). Control of T(H)17/T(reg) balance by hypoxia-inducible factor 1. *Cell*, 146(5), 772–784. <https://doi.org/10.1016/j.cell.2011.07.033>
26. Jantsch, J., & Schödel, J. (2015). Hypoxia and hypoxia-inducible factors in myeloid cell-driven host defense and tissue homeostasis. *Immunobiology*, 220(2), 305–314. <https://doi.org/10.1016/j.imbio.2014.09.009>
27. Cui, W., Zhou, J., Dehne, N., & Brüne, B. (2015). Hypoxia induces calpain activity and degrades SMAD2 to attenuate TGF β signaling in macrophages. *Cell & bioscience*, 5, 36. <https://doi.org/10.1186/s13578-015-0026-x>
28. Guzy, R. D., Hoyos, B., Robin, E., Chen, H., Liu, L., Mansfield, K. D., Simon, M. C., Hammerling, U., & Schumacker, P. T. (2005). Mitochondrial complex III is required for hypoxia-induced ROS production and cellular oxygen sensing. *Cell metabolism*, 1(6), 401–408. <https://doi.org/10.1016/j.cmet.2005.05.001>
29. Brunelle, J. K., Bell, E. L., Quesada, N. M., Vercauteren, K., Tiranti, V., Zeviani, M., Scarpulla, R. C., & Chandel, N. S. (2005). Oxygen sensing requires mitochondrial ROS but not oxidative phosphorylation. *Cell metabolism*, 1(6), 409–414. <https://doi.org/10.1016/j.cmet.2005.05.002>
30. Kapitsinou, P. P., & Haase, V. H. (2008). The VHL tumor suppressor and HIF: insights from genetic studies in mice. *Cell death and differentiation*, 15(4), 650–659. <https://doi.org/10.1038/sj.cdd.4402313>
31. Smith, T. G., Robbins, P. A., & Ratcliffe, P. J. (2008). The human side of hypoxia-inducible factor. *British journal of haematology*, 141(3), 325–334. <https://doi.org/10.1111/j.1365-2141.2008.07029.x>
32. Lee, J. W., Bae, S. H., Jeong, J. W., Kim, S. H., & Kim, K. W. (2004). Hypoxia-inducible factor (HIF-1) α : its protein stability and biological functions. *Experimental & molecular medicine*, 36(1), 1–12. <https://doi.org/10.1038/emm.2004.1>
33. Vaupel, P., & Mayer, A. (2007). Hypoxia in cancer: significance and impact on clinical outcome. *Cancer metastasis reviews*, 26(2), 225–239. <https://doi.org/10.1007/s10555-007-9055-1>